

Potential applications and design of protein delivery systems Francesco Cellesi

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Nanosized systems for protein delivery



Liposomal nanocarriers



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Thermosensitive liposomes

Liposome	Lipid composition (mol/mol)	Thermo-sensitivity	T _m (°C)	$d_{h}\left(nm\right) ^{a}$	PDI ^a	ζ -potential (mV) ^a
NTSL TSL1 TSL2 TTSL LTSL	DSPC/Chol/DSPE-PEG(2000) = 65/30/5 DPPC/DSPE-PEG(2000) = 95/5 DPPC/DSPC/DSPE-PEG2000 = 80/15/5 DPPC/DSPC/Chol/DSPE-PEG(2000) = 50/25/15/3 DPPC/Lyso-PC/DSPE-PEG2000 = 90/10/4	No YES YES YES YES	n/a [*] 42.5 43.0 40.9 40.9	$\begin{array}{c} 155\pm 2 \\ 152\pm 1 \\ 134\pm 1 \\ 147\pm 2 \\ 148\pm 1 \end{array}$	$\begin{array}{c} 0.05 \pm 0.02 \\ 0.08 \pm 0.01 \\ 0.05 \pm 0.02 \\ 0.03 \pm 0.01 \\ 0.08 \pm 0.02 \end{array}$	$\begin{array}{c} -2.1\pm 0.4\\ -2.2\pm 0.6\\ -5.4\pm 0.2\\ -1.8\pm 0.2\\ -2.0\pm 0.2\end{array}$



A) Temperature dependent release of CF from liposomes (lipid concentration 5 mM) in PBS (10 mM, pH= 7.4). The samples were incubated at the desired temperature (37-45 $^{\circ}$ C) for 5 min. The CF release was measured for each formulation. B) Time dependent CF release from LTSL at different temperature (37-42 $^{\circ}$ C). Results are given as mean of three measurements, ±SD.

BDNF - Thermosensitive liposomes



(a) Temperature-dependent percent release of BDNF from LTSL in 30 min (n = 3, p < 0.05 and p < 0.01). (b) Time-dependent percent release of BDNF in 30 min at 37 C and 42 C. After 30 min incubation at set temperature, further incubation at 37 C for additional 2 h and 48 h was carried out.



Morphology of podocytes stained by green phalloidin (scale bar 20 mm); (a) cells treated with ADR for 24 h; (b) cells damaged by ADR and incubated for 48 h with blank LTSL (c) cells damaged by ADR and incubated for 48 h with BDNF-loaded LTSL (BDNF concentration 200 ng/mL).

Macromolecules loading in liposomes

Liposome	Z-average size (nm) ^a	PDI ^a	ζ-potential (mV) ^a	LE (%) ^a	EE (%) ^a
TSL1	153 ± 1	$\textbf{0.08} \pm \textbf{0.01}$	-2.1 ± 0.4	0.023 ± 0.004	15.0 ± 2.7
TSL2	154±3	$\textbf{0.13}\pm\textbf{0.04}$	-6.5 ± 0.1	$\textbf{0.038} \pm \textbf{0.008}$	25.0 ± 5.3
TTSL	150 ± 1	0.14 ± 0.03	-0.6 ± 0.1	0.013 ± 0.001	9.0 ± 0.7
LTSL	145 ± 1	0.12 ± 0.03	-6.8 ± 0.4	0.062 ± 0.007	41.3 ± 4.7

Table 4

Table 3

Characterization of TRITC-dextran and Rho-A4PEG5 encapsulated liposomes (0.5 mM lipid suspension in PBS buffer 10 mM, pH = 7.4, 25 °C).^a Mean ± SEM, N = 3.

Characterization of FITC-albumin encapsulated liposomes (0.5 mM lipid suspension in PBS 10 mM, pH = 7.4, 25 °C).^a Mean ± SEM, N = 3.

Liposome	Payload	Z-average (nm) ^a	PDI ^a	ζ-potential (mV) ^a	LE (mol%) ^a	EE (%) ^a
TSL2	TRITC-dextran Rho-A4PEG5	$\begin{array}{c} 128\pm 6\\ 132\pm 2\end{array}$	$\begin{array}{c} 0.09 \pm 0.02 \\ 0.08 \pm 0.01 \end{array}$	$\begin{array}{c} -1.3 \pm 0.3 \\ -1.5 \pm 0.2 \end{array}$	$\begin{array}{c} 0.012 \pm 0.001 \\ 0.052 \pm 0.004 \end{array}$	$\begin{array}{c} 5.1 \pm 0.6 \\ 5.6 \pm 0.4 \end{array}$
TTSL	TRITC-dextran Rho-A4PEG5	$\begin{array}{c} 156\pm3\\ 150\pm2 \end{array}$	$\begin{array}{c} 0.03 \pm 0.01 \\ 0.06 \pm 0.01 \end{array}$	$\begin{array}{c} -1.0 \pm 0.3 \\ -0.9 \pm 0.3 \end{array}$	$\begin{array}{c} 0.009 \pm 0.001 \\ 0.049 \pm 0.005 \end{array}$	$\begin{array}{c} 3.7 \pm 0.7 \\ 5.3 \pm 0.5 \end{array}$
LTSL	TRITC-dextran Rho-A4PEG5	$\begin{array}{c} 134 \pm 2 \\ 133 \pm 3 \end{array}$	$\begin{array}{c} 0.08 \pm 0.01 \\ 0.09 \pm 0.01 \end{array}$	$\begin{array}{c} -1.0 \pm 0.1 \\ -1.5 \pm 0.2 \end{array}$	$\begin{array}{c} 0.015 \pm 0.001 \\ 0.063 \pm 0.005 \end{array}$	$\begin{array}{c} 6.0\pm0.7\\ 6.8\pm0.5\end{array}$

Table 5

Characteristics of FITC-lysozyme (LZ) and BDNF loaded liposomes with different formulations. ^a Mean \pm SEM, N = 3. Liposomal vesicles were diluted to 0.5 mM with PBS buffer (10 mM, pH = 7.4) and the particle size, size distribution and ζ -potential were measured by DLS techniques at 25 °C.

Protein	Lipid formulation	Initial protein/lipids ratio (mol/mol)	Average size (nm) ^a	PDI ^a	ζ-potential (mV) ^a	LE (mol%) ^a	EE (%) ^a
LZ LZ LZ LZ LZ BDNF	TSL1 TSL2 LTSL LTSL LTSL LTSL	$\begin{array}{c} 7.1\times10^{-4}\\ 7.1\times10^{-4}\\ 7.1\times10^{-4}\\ 1.4\times10^{-3}\\ 7.1\times10^{-3}\\ 3.7\times10^{-4} \end{array}$	$134 \pm 1 136 \pm 3 133 \pm 1 138 \pm 2 134 \pm 2 142 \pm 5$	$\begin{array}{c} 0.10 \pm 0.02 \\ 0.09 \pm 0.03 \\ 0.09 \pm 0.01 \\ 0.06 \pm 0.02 \\ 0.10 \pm 0.01 \\ 0.10 \pm 0.03 \end{array}$	$\begin{array}{c} -2.02\pm 0.20\\ -2.25\pm 0.29\\ -1.96\pm 0.35\\ -1.44\pm 0.19\\ -1.40\pm 0.30\\ -1.78\pm 0.01\end{array}$	$\begin{array}{c} 0.011\pm\ 0.006\\ 0.011\pm\ 0.003\\ 0.010\pm\ 0.001\\ 0.030\pm\ 0.001\\ 0.069\pm\ 0.005\\ 0.004\pm\ 0.001 \end{array}$	$\begin{array}{c} 15.4 \pm 0.8 \\ 15.1 \pm 0.4 \\ 14.4 \pm 0.2 \\ 21.2 \pm 0.6 \\ 9.6 \pm 0.7 \\ 11.9 \pm 0.1 \end{array}$

SiO₂ gel nanoparticles



- Non toxic with acceptable in vivo biocompatibility.
- No swelling and no change in porosity in a physiologically acceptable range of temperature and pH.
- Preparation conditions based on **mild sol-gel synthesis** that can retain the activity of entrapped enzymes



Enzyme encapsulation in SiO₂ NPs

Synthesis performed in W/O microemulsion



time (min)

Synthesis performed in buffer (SiO₂ nucleation in

presence of a positively charged enzyme)

F.Cellesi, N.Tirelli, Colloids and Surfaces A (288), 2006, 52-61

P. De leonardis et Al., Colloids and Surfaces A (580) 2019, 123734

Surface polymerisation



Protein - polymer conjugates



Grafting-from approach via ARGET ATRP



Protein-polymer hybrids, controlled MW and architecture

Immunoactive PGMA





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Mannose-PGMA Uptake



Fluorinated hyperbranched polyglycerols





W. Celentano et Al., Polymer Chemistry 2020

Contractor

Polymer Chemistry

¹⁹F-HPG for MRI and DEX delivery



Complex copolymers for proteinase rich tumors



Acknowledgment













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